

Larvicidal activities of some Euro-Asiatic plants against *Culex quinquefasciatus* Say (Diptera: Culicidae)

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ABSTRACT

Methanol extracts of the aerial parts from 31 Euro-Asiatic plant fourth instar larvae species were tested for larvicidal activity against the mosquito, *Culex quinquefasciatus* Say (Diptera: Culicidae) fourth instar larvae, under laboratory conditions. The mortality from six concentrations (5, 10, 25, 50, 100 and 200 ppm) was determined and LD₅₀ was calculated. All plant extracts showed larvicidal activity in 24 h exposure tests. The methanolic extracts of plants *Otanthus maritimus* displayed the highest larvicidal activities with LD₅₀ 7 ppm, followed by *Ammi visnaga*, *Acer pseudoplatanus*, *Acer platanoides* and *Satureja hortensis* with LO₅₀ 9, 23, 28 and 28 ppm, respectively.

Keywords: *Culex quinquefasciatus*, *Otanthus maritimus*, *Ammi visnaga*, *Acer pseudoplatanus*, *Acer platanoides* and *Satureja hortensis*.

INTRODUCTION

Mosquitoes not only cause nuisance by their bites but also transmit deadly diseases like malaria, filariasis, yellow fever, dengue and Japanese encephalitis, contribute significantly to poverty and social debility in tropical countries (Jang *et al.*, 2002). *Culex quinquefasciatus* Say (Diptera: Culicidae) is a pantropical pest and urban vector of *Wuchereria bancrofti*, *Plasmodium* (avian malaria), myxomatosis, and other diseases in some parts of the world (Holder, 1999). It has been shown to be able to carry Murray Valley encephalitis (MVE) virus in laboratory studies and MVE virus has been isolated from it in northern Western Australia. *Culex quinquefasciatus* has yielded an isolate of Ross River (RR) virus during an outbreak in New Caledonia, but from a number of laboratory studies in Australia it appears to be a poor and unlikely vector of MVE, Kunjin, RR and other arboviruses. It is a poor vector of dog heartworm, and of human filariasis in more northern tropical regions (Russell, 1993).

Synthetic insecticides are today in the fore of the mosquito controlling agents. Compared to other controlling measures in the past few decades, synthetic insecticides have been used and have produced a feed back of environmental ill effect, non-target organisms being affected and most of mosquitoes species have becoming physiologically resistant to these synthetic insecticides (VCRC, 1989; Severini *et al.*, 1993). On the other hand, some mosquito species have developed high levels of resistance to microbial control agents (Rao *et al.*, 1995). These factors have created a search for ecofriendly, biodegradable and target – specific insecticides against the mosquitoes.

Plant products have been used traditionally by human communities in many parts of the world against the vector and pest species of insects (Jacobson, 1958; Pavela, 2007). Over the past 35 years, the search for plants with novel insecticidal constituents has been intensive. Among the plant families studied Asteraceae, Lamiaceae, Meliaceae,

Piperaceae, Rutaceae crude extracts or their compounds have showed toxicity (Pavela, 2006), antifeedants (Sadek, 2003; Pavela, 2004a), insect growth regulators (Pavela, 2004b; Pavela, 2005), oviposition deterrence (Dimock and Renwick, 1991; Zhao *et al.*, 1998), suppression of calling behaviour (Khan and Saxana, 1986) and reduction of fecundity and fertility (Pavela *et al.*, 2005). Such a wide variety of effects provides potential alternatives for the use of synthetic chemical insecticides. In the present investigation an attempt was made to evaluate the larvicidal efficacy of some plant extracts against *Culex quinquefasciatus*.

MATERIAL AND METHODS

Plant extracts

Fresh plant material of each of the selected species (see Table 1) was collected in 2006. Voucher specimens of all the plant species studied were deposited in herbaria of our institute. The plant material was shade-dried and powdered. The dry powder was extracted with excess of methylalcohol (500 ml of MeOH for 100g of plant powder) for 24 h. The crude extracts were filtrated and evaporated under reduced pressure in a rotary evaporator.

Test organism

The test organism *Culex quinquefasciatus* Say was reared in the laboratory conditions. (28 ± 2°C, 70 ± 5% RH, and a photo regime of 16:8 (L:D) h.) on dog biscuits and yeast powder in the 3:1 ratio. Adults were provided with 10% sucrose solution and 1-week – old chick for blood feeding.

Larvicidal activity

Mosquito larvicidal assays were carried out according to WHO standard procedures (1996), with slight modifications. The extracts were diluted in dimethyl sulphoxide (DMSO) to prepare a serial dilution of test dosage. Early fourth instar larvae of *C. quinquefasciata* were selected and transferred in 25 ml of distilled water.

Table 1. Plants selected for the screening of larvicidal activity against *Culex quinquefasciata*

| Species | Family assayed | Plant part | Yield (%) references | Voucher | Origin |
|---|----------------|------------|----------------------|---------|---------------------------------|
| <i>Acer campestre</i> L. | Aceraceae | Leaves | 8.7 | 0760 | Prague, Czech Republic |
| <i>Acer cissifolium</i> (Siebold & Zuccarini) Koch | Aceraceae | Leaves | 8.3 | 0762 | Prague, Czech Republic |
| <i>Acer negundo</i> L. | Aceraceae | Leaves | 9.5 | 0763 | Prague, Czech Republic |
| <i>Acer platanoides</i> L. | Aceraceae | Leaves | 9.2 | 0761 | Prague, Czech Republic |
| <i>Acer pseudoplatanus</i> L. | Aceraceae | Leaves | 7.6 | 0759 | Prague, Czech Republic |
| <i>Achillea millefolium</i> L. | Asteraceae | Stem | 8.9 | 0740 | Prague, Czech Republic |
| <i>Ajuga reptans</i> L. | Lamiaceae | Stem | 10.2 | 0722 | Prague, Czech Republic |
| <i>Ammi visnaga</i> (L.) LAM. | Apiaceae | Seeds | 19.3 | 0732 | Prague, Czech Republic |
| <i>Circium arvense</i> (Savi) Ten. | Asteraceae | Stem | 5.8 | 0750 | Prague, Czech Republic |
| <i>Fraxinus exelsior</i> L. | Oleaceae | Leaves | 5.3 | 0765 | Prague, Czech Republic |
| <i>Glebionis coronarium</i> (L.) Tzvelev. Russia | Asteraceae | Flower | 9.2 | 0758 | Krasnodarskiy region, |
| <i>Humulus japonicus</i> Sieb. & Zucc. | Cannabidaceae | Leaves | 7.9 | 0725 | Prague, Czech Republic |
| <i>Hysopus officinalis</i> L. | Lamiaceae | Stem | 8.7 | 0748 | Prague, Czech Republic |
| <i>Laburnum anagyroides</i> Medik. | Leguminosae | Leaves | 15.0 | 0734 | Prague, Czech Republic |
| <i>Lavandula officinalis</i> L. | Lamiaceae | Flower | 3.1 | 0227 | Prague, Czech Republic |
| <i>Limonium. Bonduelii</i> . O. Kuntze | Plumbaginaceae | Stem | 2.8 | 0746 | Prague, Czech Republic |
| <i>Matricaria maritima</i> L. | Asteraceae | Flower | 6.8 | 0766 | Prague, Czech Republic |
| <i>Matthiola tricuspidata</i> (L.) R. Brown Russia | Brassicaceae | Stem | 7.2 | 0754 | Krasnodarskiy region, |
| <i>Melisa officinalis</i> L. | Lamiaceae | Stem | 5.7 | 0742 | Prague, Czech Republic |
| <i>Ocimum basilicum</i> L. | Lamiaceae | Stem | 7.6 | 0753 | Prague, Czech Republic |
| <i>Origanum vulgare</i> L. | Lamiaceae | Stem | 7.4 | 0729 | Prague, Czech Republic |
| <i>Otanthus maritimus</i> (L.) Hoffmanns & Link Russia | Asteraceae | Stem | 5.6 | 0755 | Krasnodarskiy region, |
| <i>Salvia farinacea</i> Benth. | Lamiaceae | Stem | 5.4 | 0713 | Prague, Czech Republic |
| <i>Salvia viridis</i> L. | Lamiaceae | Flower | 4.7 | 0731 | Prague, Czech Republic |
| <i>Salvia officinalis</i> L. | Lamiaceae | Stem | 11.3 | 0721 | Prague, Czech Republic |
| <i>Satureja hortensis</i> L. | Lamiaceae | Stem | 7.7 | 0717 | Prague, Czech Republic |
| <i>Satureja nervosa</i> Desf . | Lamiaceae | Stem | 4.2 | 0756 | Crete, Greece |
| <i>Sonchus arvensis</i> L. | Asteraceae | Stem | 5.8 | 0744 | Prague, Czech Republic |
| <i>Tanacetum vulgare</i> L. | Asteraceae | Flower | 6.1 | 0736 | Krasnodarskiy region, Russia |
| <i>Teuricum capitatum</i> L. | Lamiaceae | Stem | 5.8 | 0757 | Crete, Greece |
| <i>Thymus vulgaris</i> L. | Lamiaceae | Stem | 5.0 | 0718 | Prague, Czech Republic |

For experimental treatment, 1 ml of serial dilutions was added to 224 ml of distilled water in a 500-ml glass bowl and shaken lightly to ensure a homogenous test solution. The selected larvae were transferred in distilled water into a bowl of prepared test solution with final surface area 125 cm² (25 larvae/beaker). Four replicates were of maintained for all the five dosages (5, 10, 25, 50, 100 and 200 ppm) separately. The assays were placed in a growth chamber (L16:D9, 26 °C). The mortality was determined after 24 h of exposure, during which time no food was offered to the larvae. The control mortality was corrected

by Abbott's formula (Abbott, 1925) and LD₅₀, LD₉₀, regression equation and the 95% confidence limit was calculated by using probit analysis (Finney, 1971).

RESULTS AND DISCUSSION

The results of larvicidal activity of plant extracts are presented in Table 2. All plant extracts showed larvicidal activity in 24 h exposure tests. The methanolic extracts of plants *Otanthus maritimus* displayed the highest larvicidal activities with LD₅₀ 7 ppm, followed by *Ammi visnaga*, *Acer pseudoplatanus*, *Acer platanoides* and *Satureja hortensis* with LD₅₀ 9, 23, 28 and 28 ppm, respectively.

Table 2. Insecticidal activity of plant extracts against fourth stage larvae of *Culex quinquefasciata*

| Species | Average mortality ^a (%) ±SE | LD ₅₀ (CI95) ^b | LD ₉₀ (CI95) |
|--|---|--------------------------------------|-------------------------|
| <i>Acer campestre</i> L. | 100.0±0.0 | 52 (48-75) | 86(82-98) |
| <i>Acer cissifolium</i> (Siebold & Zuccarini) Koch | 100.0±0.0 | 70 (62-84) | 123(110-132) |
| <i>Acer negundo</i> L. | 100.0±0.0 | 51(45-63) | 89(80-93) |
| <i>Acer platanoides</i> L. | 100.0±0.0 | 28(25-39) | 62(54-73) |
| <i>Acer pseudoplatanus</i> L. | 100.0±0.0 | 23(18-33) | 76(60-89) |
| <i>Achillea millefolium</i> L. | 100.0±0.0 | 120(115-126) | 159(142-169) |
| <i>Ajuga reptans</i> L. | 46.0±3.8 | >200 | >200 |
| <i>Ammi visnaga</i> (L.) LAM. | 100.0±0.0 | 9(8-18) | 45(38-52) |
| <i>Cirsium arvense</i> (Savi) Ten. | 44.6±3.8 | >200 | >200 |
| <i>Fraxinus exelsior</i> L. | 21.0±3.2 | >200 | >200 |
| <i>Glebionis coronarium</i> (L.) Tzvelev. | 100.0±0.0 | 53(46-58) | 110(102-123) |
| <i>Humulus japonicus</i> Sieb. & Zucc. | 100.0±0.0 | 25 (21-32) | 87 (76-110) |
| <i>Hysopus officinalis</i> L. | 92.7±5.9 | 150 (140-163) | >200 |
| <i>Laburnum anagyroides</i> Medik. | 98.0±8.1 | 111 (103- 126) | 198 (156-220) |
| <i>Lavandula officinalis</i> L. | 100.0±0.0 | 59(57-72) | 123(118-128) |
| <i>Limonium. Bonduelii</i> . O. Kuntze | 52.2±12.3 | >200 | >200 |
| <i>Matricaria maritima</i> L. | 100.0±0.0 | 72(60-90) | 139(126-142) |
| <i>Matthiola tricuspidata</i> (L.) R. Brown | 9.0±3.3 | >200 | >200 |
| <i>Melisa officinalis</i> L. | 44.5±6.5 | >200 | >200 |
| <i>Ocimum basilicum</i> L. | 100.0±0.0 | 32(29-45) | 69(53-82) |
| <i>Origanum vulgare</i> L. | 88.6±7.2 | 156 (148-172) | >200 |
| <i>Otanthus maritimus</i> (L.) Hoffmanns & Link | 100.0±0.0 | 7.3(6.5-9.3) | 15(12-18) |
| <i>Salvia farinacea</i> Benth. | 68.3±3.3 | 195(183-199) | >200 |
| <i>Salvia viridis</i> L. | 100.0±0.0 | 110(105-132) | 183(153-191) |
| <i>Salvia officinalis</i> L. | 84.6±10.2 | 159(133-168) | >200 |
| <i>Satureja hortensis</i> L. | 100.0±0.0 | 28(22-37) | 56(48-72) |
| <i>Satureja nervosa</i> Desf . | 32.5±8.5 | >200 | >200 |
| <i>Sonchus arvensis</i> L. | 100.0±0.0 | 68(55-78) | 118(110-132) |
| <i>Tanacetum vulgare</i> L. | 92.1±4.4 | 178(158-188) | 198(186-201) |
| <i>Teuricum capitatum</i> L. | 48.8±12.9 | >200 | >200 |
| <i>Thymus vulgaris</i> L. | 100.0±0.0 | 48.2(39-82) | 98(79-102) |
| control | 0.0±0.0 | | |

^a The average mortality (± standard error) established in concentration 200 ppm as a maximal tested concentration.

^b The lethal concentrations with the corresponding 95% confidence intervals are shown in are shown in parenthesis.

Today, the environmental safety of an insecticide is considered to be of paramount importance. Phytochemicals may serve as suitable alternatives to synthetic insecticides in future as they are relatively safe, inexpensive, and are readily available in many areas of the world (Pavela, 2007). The screening of locally available plants for mosquito control would generate local employment, reduce dependence on expensive imported products and stimulate local efforts to enhance public health.

The crude extracts of the five plant has been found to possess larvicidal activity against the mosquito *Culex quinquefasciatus*. The biological activity of this plant extracts might be due to the various compound, including phenolics, terpenoids, and alkaloids exist in plants these compounds may jointly or independently contribute to produce larvicidal and adult emergence inhibition activity against *Culex quinquefasciatus*.

The larvicidal efficacy of *O. maritimus*, *A. visnaga*, *A. pseudoplatanus*, *A. platanoides* and *S. hortensis* are

comparable to well established insecticidal plant species. Omena *et al.* (2007) studied larvicidal activities of 51 Brazilian medicinal plants against *Aedes aegypti*. Eleven of 84 extracts (obtained from different plant parts) showed significant LD₅₀ < 100 ppm and seven species showed significant LD₅₀ < 10 ppm (from genus *Annona* sp., *Derris* sp., *Erythrina* sp. and *Pterodon* sp.). Pizzarro *et al.* (1999) studied the activity of the saponine fraction of *Agave sisalana* and estimated the LD₅₀ and LD₉₀ against 3rd instar larvae of *Culex quinquefasciatus*, that were 183 and 408 ppm, respectively. These concentrations were much higher than those reported in this study, but these authors suggested its use for control of this mosquito. The leaf extracts of *O. maritimus*, *A. visnaga*, *A. pseudoplatanus*, *A. platanoides* and *S. hortensis* is superior to various neem extracts, which are reported to be effective with LD₅₀ values ranging from 55-65 ppm against mosquito larvae (Ascher and Meisner, 1989). The median lethal concentrations (LD₅₀) of various parts of *Melia azederach* ranging from 30-40 ppm against larva of *Culex pipiens* (Al-Sharook *et al.*, 1991). The effect of the various neem extracts and various parts of *Melia azederach* was slightly lower than that reported for the *C. asiatica* leaf extract. The adult emergence inhibition activity of *C. asiatica* is also comparable to different species of plant extract in different families (Muthukrishnan *et al.*, 1999; Pushpalatha and Muthukrishnan, 1999). In our study, five extracts possess remarkable larvicidal activity against mosquito *Culex quinquefasciatus*. Further investigations are needed to elucidate this activity against a wide range of all stages mosquito species and also the active ingredients of the extract responsible for larvicidal and adult emergence inhibition activity in *Culex quinquefasciatus* should be identified and utilized, if possible, in preparing a commercial product / formulation to be used as a mosquitocidal.

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